Genome-wide association study of hippocampal atrophy rate in nondemented elders

Yu Guo¹, Wei Xu¹, Jie-Qiong Li¹, Ya-Nan Ou¹, Xue-Ning Shen², Yu-Yuan Huang², Qiang Dong², Lan Tan¹, Jin-Tai Yu², on behalf of Alzheimer's Disease Neuroimaging Initiative^{*}

¹Department of Neurology, Qingdao Municipal Hospital Affiliated to Qingdao University, Qingdao, China ²Department of Neurology and Institute of Neurology, Huashan Hospital, Shanghai Medical College, Fudan University, Shanghai, China

^{*}Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at:

http://adni.loni.usc.edu/wp-content/uploads/how to apply/ADNI Acknowledgement List.pdf

Correspondence to: Jin-Tai Yu, Lan Tan; email: jintai yu@fudan.edu.cn, dr.tanlan@163.comKeywords: hippocampal atrophy rate, cognitive decline, genome-wide association studyReceived: July 25, 2019Accepted: November 8, 2019Published: November 23, 2019

Copyright: Guo et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Hippocampal atrophy rate has been correlated with cognitive decline and its genetic modifiers are still unclear. Here we firstly performed a genome-wide association study (GWAS) to identify genetic loci that regulate hippocampal atrophy rate. Six hundred and two non-Hispanic Caucasian elders without dementia were included from the Alzheimer's Disease Neuroimaging Initiative cohort. Three single nucleotide polymorphisms (SNPs) (rs4420638, rs56131196, rs157582) in the *TOMM40-APOC1* region were associated with hippocampal atrophy rate at genome-wide significance and 3 additional SNPs (in *TOMM40* and near *MIR302F* gene) reached a suggestive level of significance. Strong linkage disequilibrium between rs4420638 and rs56131196 was found. The minor allele of rs4420638 (G) and the minor allele of rs157582 (T) showed associations with lower Minimental State Examination score, higher Alzheimer Disease Assessment Scale-cognitive subscale 11 score and smaller entorhinal volume using both baseline and longitudinal measurements, as well as with accelerated cognitive decline. Moreover, rs56131196 (P = 1.96×10^{-454}) and rs157582 (P = 9.70×10^{-434}) were risk loci for Alzheimer's disease. Collectively, rs4420638, rs56131196 and rs157582 were found to be associated with hippocampal atrophy rate. Besides, they were also identified as genetic loci for cognitive decline.

INTRODUCTION

The hippocampus is a vulnerable and plastic structure buried deep in the medial temporal lobe of human body [1]. The atrophic hippocampus is often accompanied by poor memory performance, and changes in the hippocampus provide a neural substrate for cognitive impairment that may be associated with normal aging, post-traumatic stress disorder, recurrent depression, and Cushing's syndrome [2]. Hippocampal atrophy rate has been demonstrated to be closely correlated with cognitive disorders, including the most commonly reported Alzheimer's disease (AD) and some non-AD disorders, such as frontotemporal dementia (FTD) and impaired memory [3–8]. As for mechanism, hippocampal atrophy rate may relate to the deposition of amyloid-beta (A β) and tau [9, 10]. Clinically, the atrophy rate was greater in subjects with normal cognition (NC) who converted to mild cognitive impairment (MCI) or AD than in those who remained

stable; it was greater in MCI subjects who converted to AD than in those who remained stable; and it was greater in fast AD progressors than slow ones [11, 12]. Consequently, the reduction in hippocampal volumes over time may be promising in predicting individuals at high risk of developing cognitive decline, monitoring disease trajectories at early stage, and assessing treatment efficacy in clinical practice or drug trials.

Previous genome-wide screening identified novel susceptibility genes for AD using baseline hippocampal volumes as quantitative traits [13]. However, the genetic predictors of longitudinal changes in hippocampal volumes remain poorly understood. Use of quantitative traits in genome-wide association studies (GWAS) provides novel and important insights into broader trends in correlations between genes and their associated pathways [14]. Furthermore, magnetic resonance imaging (MRI) has great advantages in visualizing structural and functional brain changes, such as sufficient sensitivity, non-invasiveness, ease of availability, and good tolerance [15]. And hippocampal volumes can be reliably measured in vivo. Therefore, we conducted a GWAS with longitudinal MRI measures of hippocampal volumes to identify genetic risk factors influencing hippocampal atrophy rate in non-demented elders. These genetic contributors may be involved in cognition-related pathophysiological process.

RESULTS

Characteristics of included subjects

This study included 226 NC (111 women, 74.7 \pm 5.3 years) and 376 MCI (152 women, 72.3 \pm 7.2 years) subjects from the Alzheimer's Disease Neuroimaging Initiative (ADNI) cohort of non-Hispanic Caucasian ancestry. The summarized characteristics of included subjects were shown in Table 1. MCI group (47.5%) had a higher frequency of ϵ 4 allele within *APOE* gene than NC group (25.7%). MCI group also had bigger baseline whole brain volume (P = 0.020) and smaller baseline hippocampal volume (P < 0.001) compared to NC group (P < 0.001).

Single nucleotide polymorphisms (SNPs) associated with hippocampal atrophy rate

There were 602 individuals identified for GWAS. After adjusting for age, gender, years of education, intracranial volume (ICV), MRI and the first three multidimensional scaling (MDS) components, 3 SNPs on chromosome 19, including rs4420638 in the *APOC1* gene (minor allele frequencies (MAF) = 0.1510, P = 9.32 × 10⁻⁹), rs56131196 in the *APOC1* gene (MAF = 0.1508, P = 1.10 × 10⁻⁸) and rs157582 in the *TOMM40* gene (MAF = 0.2937, P = 2.78 × 10⁻⁸), exhibited genome-wide

significant associations with hippocampal atrophy rate (Figure 1A and Table 2). Their association signals disappeared after including APOE E4 dosage as a covariate (Supplementary Figure 2). Strong linkage disequilibrium (LD) ($R^2 = 0.987$, D' = 0.993) between rs4420638 and rs56131196 was found, which was calculated using the 1,000 Genomes European cohort (Supplementary Figure 3). Accordingly, rs4420638 and rs157582 were chosen as the two top SNPs, for which the minor allele of rs4420638 (G) (AA: 0.003478 ± 0.012252, AG: -0.003415 ± 0.017455, GG: -0.003895 ± 0.013808; P $= 3.20 \times 10^{-7}$) and the minor allele of rs157582 (T) (CC: 0.003567 ± 0.012731 ; CT: -0.002717 ± 0.016733 , TT: -0.004347 ± 0.012037 ; P = 1.23×10^{-8}) were significantly associated with higher hippocampal atrophy rate in a dose-dependent manner (Figure 2).

SNPs mapped closely to the region of top SNPs were also analyzed (Figure 1B). After controlling for the genotypes of the two top SNPs (rs4420638, rs157582) (Figure 1C and 1D), no SNPs showed strong associations with hippocampal atrophy rate in this region, indicating that these nearby SNPs might be driven by the two top SNPs. The Quantile-Quantile plot didn't show evidence of spurious inflation in test statistics (the genomic inflation factor = 1) due to population stratification or other confounders (Supplementary Figure 4).

Three SNPs were associated with hippocampal atrophy rate at suggestive levels of significance ($P < 1 \times 10^{-5}$) (Figure 1A and Supplementary Table 1). These loci comprised SNPs within the *TOMM40* gene (rs2075650) as well as SNPs near the *MIR302F* gene (rs4271662 and rs2900721). The association signals in these SNPs disappeared when *APOE* ε 4 dosage was included as a covariate (Supplementary Figure 2 and Supplementary Table 2).

Impact of rs4420638 and rs157582 on cognitive scores and brain structures

Associations of two top SNPs with cognitive scores and brain structures were analyzed. In the crosssectional analyses, both the minor allele of rs4420638 (G) and the minor allele of rs157582 (T) were associated with lower Mini-mental State Examination (MMSE) score, higher Alzheimer Disease Assessment Scale-cognitive subscale 11 (ADAS-cog 11) score and smaller entorhinal volume (Supplementary Figure 5). In the longitudinal study, subjects with a minor allele of rs4420638 (G) showed faster cognitive decline in MMSE score (P < 0.0001) and ADAS-cog 11 score (P < 0.0001), as well as greater rates of entorhinal atrophy (P = 0.0001) and ventricular enlargement (P < 0.0001). Besides, subjects with a minor allele of rs157582 (T) also showed faster cognitive decline in

Table 1. Demographic information.

Baseline diagnosis	NC	MCI	Total
Sample size, n (%)	226 (37.5)	376 (62.5)	602
Age at baseline, mean (SD)	74.7 (5.3)	73.2 (7.2)	73.2 (6.7)
Females, n (%)	111 (49.1)	152 (40.3)	263 (43.6)
Education years, mean (SD)	16.4 (2.7)	15.9 (2.9)	16.1 (2.8)
APOE ε4 carrier, n (%)	58 (25.7)	178 (47.5)	236 (39.7)
MMSE at baseline, mean (SD)	29.1 (1.1)	27.9 (1.6)	28.4 (1.6)
WBV at baseline, mean (SD)	1,033,459.2 (100,326.4)	1,053,883.0 (108,862.1)	1,046,258.1 (106,216.4)
HPV at baseline, mean (SD) ^a	7,296.7 (881.2)	6,950.0 (1,124.4)	7,080.1 (1,053.3)

Abbreviations: *APOE, apolipoprotein E*; HPV, hippocampal volume; MCI, mild cognitive impairment; MMSE, Mini-mental State Examination; NC, normal cognition; SD, standard deviation; WBV, whole brain volume.

^aThe MCI group had lower baseline hippocampal volumes compared to the NC group (P < 0.001).



Figure 1. Manhattan and regional plots for associations with hippocampal atrophy rate. (A) Genome-wide signal intensity (Manhattan) plots showing the $-\log_{10}$ (p value) for individual single nucleotide polymorphisms. (B) Regional association results for the 45.0 Mb to 45.8 Mb region of chromosome 19. (C) Association results for the 45.0 Mb to 45.8 Mb region of chromosome 19 controlling for rs4420638. (D) Association results for the 45.0 Mb to 45.8 Mb region of chromosome 19 controlling for rs157582.

Table 2. Top SNPs associated with hippocampal atrophy rate.

SNP	CHR	BP	MAF	Closest Gene	SNP Type	BETA	Р
rs4420638	19	45422946	G=0.1510	APOC1	intron	-0.005454	9.32E-09
rs56131196	19	45422846	A=0.1508	APOC1	intron	-0.005557	1.10E-08
rs157582	19	45396219	T=0.2937	TOMM40	intron	-0.005351	2.78E-08

Abbreviations: BP, base pair (variant position); CHR, chromosome; MAF, minor allele frequency; SNP, single nucleotide polymorphism.

MMSE score (P < 0.0001) and ADAS-cog 11 score (P < 0.0001), as well as greater rates of entorhinal atrophy (P = 0.0049) and ventricular enlargement (P < 0.0001) (Figure 3).



Figure 2. Hippocampal atrophy rates of different genotypes. The y-axis showed the hippocampal atrophy rate and the x-axis corresponded to different genotypes. The effect of genotypes on hippocampal atrophy rate was examined with a multiple linear regression model using age, gender and diagnosis as covariates. (A) The minor allele of rs4420638 (G) showed association with hippocampal atrophy rate in a dose-dependent manner (P = 3.20×10^{-7}). (B) The minor allele of rs157582 (T) showed association with hippocampal atrophy rate in a dose-dependent manner (P = 1.23×10^{-8}).

Effect of SNPs on risk of cognitive decline

After the GWAS, we conducted survival analysis to further explore the influence of two top SNPs on cognitive decline (Figure 4 and Supplementary Table 3). The COX regression analysis was performed on minor allele homozygotes, heterozygotes and major allele homozygotes by including age and gender as covariates. Both the minor allele of rs4420638 (G) (P = 2.04×10^{-12}) and the minor allele of rs157582 (T) (P = 8.23×10^{-8}) appeared to accelerate cognitive decline, conferring increased risk to homozygous and heterozygous carriers of the minor allele and confirming the positive direction of effects detected in our GWAS. The GG (hazard ratio (HR) = 3.917, 95% confidence interval (CI) = 2.478 to 6.191) and AG (HR = 2.212, 95% CI = 1.615 to 3.029) genotypes of rs4420638 showed a greater risk of cognitive decline than the AA genotype. Also, the TT (HR = 4.190, 95% CI = 2.502 to 7.016) and CT (HR = 1.704, 95% CI = 1.201 to 2.418) genotypes of rs157582 showed a greater risk of cognitive decline than the CC genotype.

Bioinformatics analyses

The International Genomics of Alzheimer's Disease Project (IGAP) is by far the largest genetic epidemiological survey of AD risk, which was performed in two stages comprising a discovery step (stage 1) and a replication step (stage 2). Based on several grand-scale meta-analyses, the IGAP has identified many susceptibility loci for AD [16, 17]. After checking the top loci linked to hippocampal atrophy rate in the stage 1 meta-analysis from IGAP database, we identified rs56131196 (P = 1.96×10^{-454}) and rs157582 (P = 9.70×10^{-434}) as risk loci for AD.

As for expression quantitative trait loci (eQTL) analyses, the minor allele of rs4420638 (G) or the minor allele of rs56131196 (A) was upregulated in frontal cortex (P = 0.02) (Supplementary Figure 6). In accordance with the Allen Institute Human Brain Atlas, *APOC1* appeared to be selectively expressed in the hippocampus region. *APOC1* could also be regulated in the temporal and visual cortices (http://human.brain-

map.org/). Besides, rs157582 has a modest cis-eQTL effect on *GEMIN7* gene (Z = 3.03, P = 2.41×10^{-3}) in whole blood (Supplementary Figure 6) [18].

DISCUSSION

This study was the first to conduct a GWAS of hippocampal atrophy rate in non-demented elders. We identified genome-wide significant associations of 3 SNPs (rs4420638, rs56131196, rs157582) in the *TOMM40-APOC1* region with hippocampal atrophy rate and 3

additional suggestive association loci (in *TOMM40* gene and near *MIR302F* gene). The minor allele of rs4420638 (G) and the minor allele of rs157582 (T) showed associations with lower MMSE score, higher ADAS-cog 11 score and smaller entorhinal volume using both baseline and longitudinal measurements, as well as with accelerated cognitive decline. Our findings provide evidence that *TOMM40* and *APOC1* as candidate genes may promote the application of hippocampal atrophy rate as an early biomarker for predicting cognitive progression and detecting disease trajectories.



Figure 3. Impact of rs4420638 and rs157582 on longitudinal measurements in cognitive scores and brain structures. Associations of rs4420638 with longitudinal measurements in Mini-mental State Examination (MMSE) score (A), Alzheimer Disease Assessment Scale-cognitive subscale 11 (ADAS-cog 11) score (B), entorhinal volume (C) and ventricular volume (D) over time. Associations of rs157582 with longitudinal measurements in MMSE score (E), ADAS-cog 11 score (F), entorhinal volume (G) and ventricular volume (H) over time.



Figure 4. Kaplan–Meier survival curves for probability of cognitive progression. Numbers of individuals at risk at different followup time points were presented. Survival time was calculated according to the interval from the initial baseline evaluation to cognitive progression. (A) Subjects with the minor allele of rs4420638 (G) showed a greater rate of cognitive decline. (B) Subjects with the minor allele of rs157582 (T) showed a greater rate of cognitive decline.

Both APOC1 and TOMM40 are genes adjacent to APOE, which showed no significant associations with hippocampal atrophy rate after adjusting for APOE E4 dosage. Although the most concise explanation may be that APOE £4 was driving the observed associations, the extensive LD structure around APOE and various associations reported in APOE region made it difficult to further explain the result. As in any SNP-trait association, only statistical association was not enough to infer causality [19]. Besides, it's impossible for us to analyze all potentially causal variants in the region. A previous study also supported that the correlation between APOE region and cognitive function was not mediated solely by the APOE ɛ4 allele [20]. And the polymorphic poly-T variant in the TOMM40 gene provided among different populations more possibilities for the interpretation of our results [21]. Compared with those having shorter poly-T repeats, APOE $\varepsilon 3$ carriers having a long poly-T repeat in the TOMM40 gene not only have an earlier age of onset of late-onset AD, but also have decreased memory abilities and grey matter volumes [20]. It seems that the polymorphic poly-T variant in the TOMM40 gene provides greatly improved accuracy in the estimation of cognitive disorders for APOE ɛ3 carriers. Furthermore, a large-scale longitudinal study reported that both APOE ɛ4 and poly-T repeats in the TOMM40 gene were associated with cognitive decline, but there was no interaction between the two genes [22]. Thus, it's difficult to say whether the correlations between TOMM40-APOE-APOC1 region and cognitive function were mediated by the APOE ɛ4 allele or by poly-T repeats, or were mediated by more complex mechanisms. More research is warranted to explore the related pathogenesis.

Rs56131196 and rs157582 have been reported in GWAS studies on AD and aging-related verbal memory respectively [23, 24], thus lending validation and confidence to our analytic procedure and results. Previous studies suggested that the elderly carrying APOC1 gene tended to perform worse in cognitive showed more severe hippocampal scores and abnormalities [25, 26]. Besides, the APOCI-knock-out mice might have worse memory performance than those carrying APOC1 [27]. As for mechanism, ApoC1 (apolipoprotein C1) encoded by APOC1 is a member of the apolipoprotein family, which may be involved in multiple biological processes comprising cholesterol metabolism and neuronal apoptosis [28]. But the specific mechanisms by which APOC1 gene modulates the risk of cognitive impairment remain controversial, although some research has been done in this field [26, 29, 30]. The protein that TOMM40 encodes, TOM40 (translocase of the outer mitochondrial membrane 40), constitutes an external mitochondrial membrane channel that promotes the transport of many aggregating proteins to mitochondria [31, 32]. TOM40 protein may also act as a molecular chaperone that could accelerate the movement of ribosomal preproteins through the channel and assemble them in the mitochondria after translation [31, 32]. By mediating the dynamic functions of mitochondria, *TOMM40* may contribute to changes in cognitive status.

Both TOMM40 and APOC1 are in strong LD with APOE on chromosome 19. Several SNPs in TOMM40-APOE-APOC1 region have been detected to be associated with cognitive impairment [19, 33]. Each of these 3 adjacent genes could encode proteins with biological values that may affect cognitive function. However, controversy still exists as to whether the associations between these 3 adjacent genes and cognition are independent of APOE allele or are driven by LD with APOE. The biochemical interaction between APOC1 and APOE may be associated with cognitive impairment, since the binding of triglyceride lipoproteins to the very low density lipoprotein receptor, mediated by APOE allele, could be interfered by overexpressed ApoC1 [26]. Furthermore, APOC1 could increase the risk of cognitive impairment by modulating lipid metabolism. Additionally, apolipoprotein E (ApoE), amyloid, and synuclein proteins are interactive with Tom40 [34]. The Tom40 protein forms the channel through which amyloid beta protein precursor (APP) and AB travel and aggregate to cause mitochondrial abnormalities [32]. There was also evidence for the effects of APOE receptors on APP trafficking and AB production, and the effects of *APOE* on A β aggregation and clearance [35]. Thus, it has been postulated that APOE and TOMM40 genes might share similar mechanisms in mediating disease risk [32].

A functional analysis targeting TOMM40-APOE-APOC1 region reported that various APOE locus cisregulatory elements affect both APOE and TOMM40 promoter activity [36]. This indicates that gene expression patterns in this region may be modulated by a complicated transcriptional regulatory structure. Evidence also supported the role of epigenetic mechanisms such as deoxyribonucleic acid (DNA) methylation in the regulation of gene expression [19]. The increase of DNA methylation often leads to downregulated gene expression by either blocking access of transcriptional factors or enrolling methyl-cytosineguanine-binding proteins [37]. The repressed gene expression within the region was demonstrated to be correlated with cognitive dysfunction both in blood samples and brain tissues [38-40]. Both the methylation-gene expression and gene expressioncognition associations in the TOMM40-APOE-APOC1 region are worth investigating in the future.

There was a suggestive finding in the *MIR302F* gene, which was a member of microRNA (micro ribonucleic acid) family that participated in post-transcriptional regulation of gene expression in multicellular organisms via influencing both the stability and translation of messenger ribonucleic acids [41]. Studies have reported the associations of breast cancer [42], gastric cancer [42, 43] and acute heart failure [44] with *MIR302F* gene, whereas little was known about the mechanism by which *MIR302F* gene correlated with cognitive deterioration. Further investigations are especially warranted to explore how *MIR302F* gene influences the progression of cognitive disorders.

Limitations

Some limitations must also be acknowledged. Firstly, the sample size was relatively small, which may not be representative of the general population. Secondly, our participants were restricted to non-Hispanic Caucasians and we didn't explore the diversity among different populations. Thirdly, we applied Bonferroni correction for multiple comparisons and set the MAF threshold at > 0.10, which could enhance statistical power to avoid false-positive results but may miss less common SNPs.

METHODS

ADNI dataset

The ADNI was launched in 2003 as a public-private partnership, led by Principal Investigator Michael W. Weiner, MD, VA Medical Center and University of California–San Francisco. The primary goal of ADNI has been to investigate the effectiveness of integrating neuroimaging, genetic/biological markers, as well as clinical and neuropsychological assessments in measuring the progression of MCI and early AD. All ADNI individuals were recruited from over 50 sites across the United States and Canada, and most people were non-Hispanic Caucasians.

Participants

In this study, 602 non-Hispanic Caucasian nondemented individuals (NC = 226, MCI = 376) were enrolled from the ADNI cohort after applying quality control (QC) procedures. All participants received baseline and periodic clinical and neuropsychological assessments as well as serial MRI. Data used in the preparation for this article were derived from the ADNI database (<u>http://adni.loni.usc.edu/</u>).

A total of 698 samples before QC were available with both GWAS data and hippocampus data. To reduce confounding effects by genetic ancestry that could lead to population stratification, the analysis data was restricted to non-Hispanic Caucasian participants (n = 648). To avoid the impact of AD pathology on results, all participants were restricted to cognitively normal individuals or those with mild cognitive impairment (n = 610). To determine cryptically related individuals and/or sample mix-ups, identify-by-descent estimates and MDS components were conducted using PLINK [45]. This step excluded 5 participants who showed cryptically associated and clustering separately from the other subjects (Supplementary Figure 1), remaining 605 valid samples. Finally, all samples presented tight clustering with the population of European descent using the HapMap cohort.

Individuals were followed up to detect progressive cognitive decline defined as (1) losing > 3 points between the first and last MMSE measurements, (2) developing from NC to MCI or from MCI to dementia, or (3) having a score < 24 at last MMSE [46].

Hippocampal atrophy rate and QC

Longitudinal hippocampal volume measurements by MRI could be downloaded from the ADNI database, which was conducted by N. Schuff and his colleagues at UCSF via FreeSurfer version 4.3 [47]. The hippocampal atrophy rates were obtained from a mixed-effects model using "arm, lme4 and lmerTest" packages in R software, after controlling for age of entry, gender, number of APOE E4 allele, years of education, baseline diagnosis and total ICV. Individualized rate was then used as a quantitative outcome phenotype for the GWAS. QC was conducted to mitigate the impact of extreme values on statistical results. The mean and standard deviation (SD) of the hippocampal atrophy rate were calculated by experienced operators blinded to clinical data, and the figures greater or smaller than 4fold SD from the mean value were considered as extreme outliers and removed from this analysis. After eliminating 3 outliers, there were 602 valid subjects left.

Genotyping and QC

Genotyping for all samples was analyzed by the Illumina Human Hap610-Quad BeadChips featuring 2,379,855 SNPs. QC procedures were implemented with the PLINK software following the stringent criteria: call rates for SNPs > 98%, call rates for individuals > 95%, MAF > 0.10 and Hardy-Weinberg equilibrium test P > 0.001. We restricted the MAF value > 0.10 for SNPs to avoid potentially false-positive results and improve statistical power. Finally, all 602 subjects remained in the analysis and 695,203 SNPs passed QC protocols. The overall genotyping rate in remaining individuals was 99.7%.

Statistical analyses

One-way analysis of variance (ANOVA) and Turkey's multiple comparison tests were used to determine the difference in baseline hippocampal volumes among different diagnostic groups. The associations of hippocampal atrophy rate with genetic polymorphisms were determined using multiple linear regression under an additive genetic model in PLINK v1.9 software. To correct for confounding due to population stratification, the first three MDS components were calculated in PLINK and applied as covariates in the regression model. Age of entry, gender, years of education, ICV and MRI were also included as covariates. To investigate the effect of APOE £4 on hippocampal atrophy rate, the above GWAS analyses were repeated with APOE E4 dosage fitted as a covariate. To account for multiple comparisons, Bonferroni correction was applied and thresholds of P < 5 \times 10⁻⁸ and P < 1 \times 10⁻⁵ were used for genome-wide significant and suggestive associations, respectively [48]. Genome-wide associations were visualized using R package "qqman" and regional association plots were generated with the LocusZoom web tool (http://locuszoom.org/). LD analysis was performed using HaploReg v4.1 based on data from the 1000 Genomes Project (EUR). The difference in hippocampal atrophy rates among different genotype groups was determined using a multiple linear regression model in R software after adjusting for age, gender and diagnosis.

R software was also applied to explore the correlations of top SNPs with cognitive scores (MMSE score and ADAS-cog 11 score) and specific brain structures (entorhinal volume and ventricular volume) after adjusting for age, gender, years of education, ICV and MRI. The associations of top SNPs with the above indexes from both cross-sectional and longitudinal perspectives were determined using the multiple linear regression and linear mixed models, respectively. In the survival analysis, Kaplan-Meier survival curves were used to present trajectories of cognitive progression and COX regression model with age and gender as covariates was used to investigate the influence of two top SNPs on cognitive decline.

Bioinformatics analyses

SNP annotations were performed using the NCBI Database (http://www.ncbi.nlm.nih.gov/SNP/). The IGAP was searched to determine the associations of top SNPs with AD risk. EQTL analyses were conducted using multiple publicly available datasets in human brain tissues (http://BRAINEAC.org; Allen Institute Human Brain Atlas; http://human.brain-map.org/) and the whole blood (http://www.genenetwork. nl/bloodeqtlbrowser/).

CONCLUSIONS

In summary, we detected 3 genome-wide significant SNPs (rs4420638, rs56131196, rs157582) in the *TOMM40-APOC1* region and 3 suggestive loci (in *TOMM40* and near *MIR302F*) associated with hippocampal atrophy rate among non-demented elders. Since the 3 top SNPs were also identified as genetic loci for cognitive decline, our study indicated that hippocampal atrophy rate may be promising for monitoring cognitive progression.

CONFLICTS OF INTEREST

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

FUNDING

This study was supported by grants from the National Key R&D Program of China (2016YFC1305803), the National Natural Science Foundation of China (91849126, 81571245, and 81771148), Shanghai Municipal Science and Technology Major Project (No.2018SHZDZX01) and ZHANGJIANG LAB, Tianqiao and Chrissy Chen Institute, and the State Key Laboratory of Neurobiology and Frontiers Center for Brain Science of Ministry of Education, Fudan University. Data collection and sharing for this project was funded by the Alzheimer's Disease Neuroimaging Initiative (ADNI) (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research and Development, LLC.; Johnson and Johnson Pharmaceutical Research and Development LLC.; Lumosity; Lundbeck; Merck and Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier: Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (http://www.fnih.org). The grantee organization is the Northern California

Institute for Research and Education, and the study is coordinated by the Alzheimer's Therapeutic Research Institute at the University of Southern California. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California.

REFERENCES

- Knierim JJ. The hippocampus. Curr Biol. 2015; 25:R1116–21. <u>https://doi.org/10.1016/j.cub.2015.10.049</u> PMID:<u>26654366</u>
- McEwen BS. Possible mechanisms for atrophy of the human hippocampus. Mol Psychiatry. 1997; 2:255–62. <u>https://doi.org/10.1038/sj.mp.4000254</u> PMID:9152991
- den Heijer T, van der Lijn F, Koudstaal PJ, Hofman A, van der Lugt A, Krestin GP, Niessen WJ, Breteler MM. A 10-year follow-up of hippocampal volume on magnetic resonance imaging in early dementia and cognitive decline. Brain. 2010; 133:1163–72. https://doi.org/10.1093/brain/awq048 PMID:20375138
- Barnes J, Bartlett JW, van de Pol LA, Loy CT, Scahill RI, Frost C, Thompson P, Fox NC. A meta-analysis of hippocampal atrophy rates in Alzheimer's disease. Neurobiol Aging. 2009; 30:1711–23. <u>https://doi.org/10.1016/j.neurobiolaging.2008.01.010</u> PMID:<u>18346820</u>
- Frankó E, Joly O, and Alzheimer's Disease Neuroimaging Initiative. Evaluating Alzheimer's disease progression using rate of regional hippocampal atrophy. PLoS One. 2013; 8:e71354. <u>https://doi.org/10.1371/journal.pone.0071354</u> PMID:<u>23951142</u>
- Henneman WJ, Sluimer JD, Barnes J, van der Flier WM, Sluimer IC, Fox NC, Scheltens P, Vrenken H, Barkhof F. Hippocampal atrophy rates in Alzheimer disease: added value over whole brain volume measures. Neurology. 2009; 72:999–1007. <u>https://doi.org/10.1212/01.wnl.0000344568.09360.31</u> PMID:<u>19289740</u>
- Whitwell JL, Weigand SD, Gunter JL, Boeve BF, Rademakers R, Baker M, Knopman DS, Wszolek ZK, Petersen RC, Jack CR Jr, Josephs KA. Trajectories of brain and hippocampal atrophy in FTD with mutations in MAPT or GRN. Neurology. 2011; 77:393–98. <u>https://doi.org/10.1212/WNL.0b013e318227047f</u> PMID:<u>21753165</u>
- Mungas D, Harvey D, Reed BR, Jagust WJ, DeCarli C, Beckett L, Mack WJ, Kramer JH, Weiner MW, Schuff N, Chui HC. Longitudinal volumetric MRI change and rate of cognitive decline. Neurology. 2005; 65:565–71.

https://doi.org/10.1212/01.wnl.0000172913.88973.0d PMID:<u>16116117</u>

 Andrews KA, Frost C, Modat M, Cardoso MJ, Rowe CC, Villemagne V, Fox NC, Ourselin S, Schott JM, Rowe CC, Villemagne V, Fox NC, Ourselin S, Schott JM, and AIBL. Acceleration of hippocampal atrophy rates in asymptomatic amyloidosis. Neurobiol Aging. 2016; 39:99–107.

https://doi.org/10.1016/j.neurobiolaging.2015.10.013 PMID:26923406

 Henneman WJ, Vrenken H, Barnes J, Sluimer IC, Verwey NA, Blankenstein MA, Klein M, Fox NC, Scheltens P, Barkhof F, van der Flier WM. Baseline CSF p-tau levels independently predict progression of hippocampal atrophy in Alzheimer disease. Neurology. 2009; 73:935–40.

https://doi.org/10.1212/WNL.0b013e3181b879ac PMID:<u>19770469</u>

- Jack CR Jr, Petersen RC, Xu Y, O'Brien PC, Smith GE, Ivnik RJ, Tangalos EG, Kokmen E. Rate of medial temporal lobe atrophy in typical aging and Alzheimer's disease. Neurology. 1998; 51:993–99. https://doi.org/10.1212/WNL.51.4.993 PMID:9781519
- Jack CR Jr, Shiung MM, Gunter JL, O'Brien PC, Weigand SD, Knopman DS, Boeve BF, Ivnik RJ, Smith GE, Cha RH, Tangalos EG, Petersen RC. Comparison of different MRI brain atrophy rate measures with clinical disease progression in AD. Neurology. 2004; 62:591–600. <u>https://doi.org/10.1212/01.WNL.0000110315.26026.EF</u> PMID:<u>14981176</u>
- Potkin SG, Guffanti G, Lakatos A, Turner JA, Kruggel F, Fallon JH, Saykin AJ, Orro A, Lupoli S, Salvi E, Weiner M, Macciardi F, and Alzheimer's Disease Neuroimaging Initiative. Hippocampal atrophy as a quantitative trait in a genome-wide association study identifying novel susceptibility genes for Alzheimer's disease. PLoS One. 2009; 4:e6501.

https://doi.org/10.1371/journal.pone.0006501 PMID:<u>19668339</u>

- Ramanan VK, Shen L, Moore JH, Saykin AJ. Pathway analysis of genomic data: concepts, methods, and prospects for future development. Trends Genet. 2012; 28:323–32. <u>https://doi.org/10.1016/j.tig.2012.03.004</u>
 PMID:22480918
- Duncan GW, Firbank MJ, O'Brien JT, Burn DJ. Magnetic resonance imaging: a biomarker for cognitive impairment in Parkinson's disease? Mov Disord. 2013; 28:425–38. <u>https://doi.org/10.1002/mds.25352</u> PMID:<u>23450518</u>
- 16. Witoelar A, Rongve A, Almdahl IS, Ulstein ID, Engvig A, White LR, Selbæk G, Stordal E, Andersen F, Brækhus A,

Saltvedt I, Engedal K, Hughes T, et al. Meta-analysis of Alzheimer's disease on 9,751 samples from Norway and IGAP study identifies four risk loci. Sci Rep. 2018; 8:18088. <u>https://doi.org/10.1038/s41598-018-36429-6</u> PMID:<u>30591712</u>

- Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, DeStafano AL, Bis JC, Beecham GW, Grenier-Boley B, Russo G, Thorton-Wells TA, Jones N, et al, and Cohorts for Heart and Aging Research in Genomic Epidemiology. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. Nat Genet. 2013; 45:1452–58. <u>https://doi.org/10.1038/ng.2802</u> PMID:<u>24162737</u>
- Westra HJ, Peters MJ, Esko T, Yaghootkar H, Schurmann C, Kettunen J, Christiansen MW, Fairfax BP, Schramm K, Powell JE, Zhernakova A, Zhernakova DV, Veldink JH, et al. Systematic identification of trans eQTLs as putative drivers of known disease associations. Nat Genet. 2013; 45:1238–43. <u>https://doi.org/10.1038/ng.2756</u> PMID:24013639
- Davies G, Harris SE, Reynolds CA, Payton A, Knight HM, Liewald DC, Lopez LM, Luciano M, Gow AJ, Corley J, Henderson R, Murray C, Pattie A, et al. A genome-wide association study implicates the APOE locus in nonpathological cognitive ageing. Mol Psychiatry. 2014; 19:76–87. <u>https://doi.org/10.1038/mp.2012.159</u> PMID:<u>23207651</u>
- Roses AD, Lutz MW, Amrine-Madsen H, Saunders AM, Crenshaw DG, Sundseth SS, Huentelman MJ, Welsh-Bohmer KA, Reiman EM. A TOMM40 variable-length polymorphism predicts the age of late-onset Alzheimer's disease. Pharmacogenomics J. 2010; 10:375–84. <u>https://doi.org/10.1038/tpj.2009.69</u> PMID:20029386
- 21. Ghebranious N, Ivacic L, Mallum J, Dokken C. Detection of ApoE E2, E3 and E4 alleles using MALDI-TOF mass spectrometry and the homogeneous mass-extend technology. Nucleic Acids Res. 2005; 33:e149. https://doi.org/10.1093/nar/gni155 PMID:16204452
- Schiepers OJ, Harris SE, Gow AJ, Pattie A, Brett CE, Starr JM, Deary IJ. APOE E4 status predicts age-related cognitive decline in the ninth decade: longitudinal follow-up of the Lothian Birth Cohort 1921. Mol Psychiatry. 2012; 17:315–24. https://doi.org/10.1038/mp.2010.137 PMID:21263443
- Gao L, Cui Z, Shen L, Ji HF. Shared Genetic Etiology between Type 2 Diabetes and Alzheimer's Disease Identified by Bioinformatics Analysis. J Alzheimers Dis. 2016; 50:13–17. PMID:26639962
- 24. Arpawong TE, Pendleton N, Mekli K, McArdle JJ, Gatz M, Armoskus C, Knowles JA, Prescott CA. Genetic

variants specific to aging-related verbal memory: insights from GWASs in a population-based cohort. PLoS One. 2017; 12:e0182448. https://doi.org/10.1371/journal.pone.0182448 PMID:28800603

 Bartrés-Faz D, Junqué C, Clemente IC, Serra-Grabulosa JM, Guardia J, López-Alomar A, Sánchez-Aldeguer J, Mercader JM, Bargalló N, Olondo M, Moral P. MRI and genetic correlates of cognitive function in elders with memory impairment. Neurobiol Aging. 2001; 22:449–59.

https://doi.org/10.1016/S0197-4580(01)00207-X PMID:<u>11378252</u>

 Bartrés-Faz D, Clemente IC, Junqué C, Valveny N, López-Alomar A, Sánchez-Aldeguer J, López-Guillén A, Moral P. APOE and APOC1 genetic polymorphisms in age-associated memory impairment. Neurogenetics. 2001; 3:215–19.

https://doi.org/10.1007/s100480100122 PMID:<u>11714102</u>

- Berbée JF, Vanmierlo T, Abildayeva K, Blokland A, Jansen PJ, Lütjohann D, Gautier T, Sijbrands E, Prickaerts J, Hadfoune M, Ramaekers FC, Kuipers F, Rensen PC, Mulder M. Apolipoprotein Cl knock-out mice display impaired memory functions. J Alzheimers Dis. 2011; 23:737–47. PMID:21157034
- Leduc V, Jasmin-Bélanger S, Poirier J. APOE and cholesterol homeostasis in Alzheimer's disease. Trends Mol Med. 2010; 16:469–77. <u>https://doi.org/10.1016/j.molmed.2010.07.008</u> PMID:<u>20817608</u>
- Drigalenko E, Poduslo S, Elston R. Interaction of the apolipoprotein E and Cl loci in predisposing to late-onset Alzheimer's disease. Neurology. 1998; 51:131–35. <u>https://doi.org/10.1212/WNL.51.1.131</u> PMID:<u>9674791</u>
- 30. Poduslo SE, Neal M, Herring K, Shelly J. The apolipoprotein CI A allele as a risk factor for Alzheimer's disease. Neurochem Res. 1998; 23:361–67. <u>https://doi.org/10.1023/A:1022409617539</u> PMID:9482248
- 31. Willette AA, Webb JL, Lutz MW, Bendlin BB, Wennberg AM, Oh JM, Roses A, Koscik RL, Hermann BP, Dowling NM, Asthana S, Johnson SC, and Alzheimer's Disease Neuroimaging Initiative. Family history and TOMM40 '523 interactive associations with memory in middleaged and Alzheimer's disease cohorts. Alzheimers Dement. 2017; 13:1217–25. <u>https://doi.org/10.1016/j.jalz.2017.03.009</u>

PMID:28549947

 Burggren AC, Mahmood Z, Harrison TM, Siddarth P, Miller KJ, Small GW, Merrill DA, Bookheimer SY. Hippocampal thinning linked to longer TOMM40 poly-T variant lengths in the absence of the APOE ε4 variant. Alzheimers Dement. 2017; 13:739–48. <u>https://doi.org/10.1016/j.jalz.2016.12.009</u> PMID:<u>28183529</u>

- Lutz MW, Crenshaw DG, Saunders AM, Roses AD. Genetic variation at a single locus and age of onset for Alzheimer's disease. Alzheimers Dement. 2010; 6:125–31. <u>https://doi.org/10.1016/j.jalz.2010.01.011</u> PMID:<u>20298972</u>
- 34. Roses A, Sundseth S, Saunders A, Gottschalk W, Burns D, Lutz M. Understanding the genetics of APOE and TOMM40 and role of mitochondrial structure and function in clinical pharmacology of Alzheimer's disease. Alzheimers Dement. 2016; 12:687–94. <u>https://doi.org/10.1016/j.jalz.2016.03.015</u> PMID:27154058
- 35. Yu JT, Tan L, Hardy J. Apolipoprotein E in Alzheimer's disease: an update. Annu Rev Neurosci. 2014; 37:79–100.
 https://doi.org/10.1146/annurev-neuro-071013-014300 PMID:24821312
- Bekris LM, Lutz F, Yu CE. Functional analysis of APOE locus genetic variation implicates regional enhancers in the regulation of both TOMM40 and APOE. J Hum Genet. 2012; 57:18–25. https://doi.org/10.1038/jhg.2011.123 PMID:22089642
- Liu J, Zhao W, Ware EB, Turner ST, Mosley TH, Smith JA. DNA methylation in the APOE genomic region is associated with cognitive function in African Americans. BMC Med Genomics. 2018; 11:43. <u>https://doi.org/10.1186/s12920-018-0363-9</u> PMID:<u>29739406</u>
- Yasuno F, Tanimukai S, Sasaki M, Hidaka S, Ikejima C, Yamashita F, Kodama C, Mizukami K, Michikawa M, Asada T. Association between cognitive function and plasma lipids of the elderly after controlling for apolipoprotein E genotype. Am J Geriatr Psychiatry. 2012; 20:574–83.

https://doi.org/10.1097/JGP.0b013e318211819b PMID:<u>21407046</u>

- Lambert JC, Mann D, Richard F, Tian J, Shi J, Thaker U, Merrot S, Harris J, Frigard B, Iwatsubo T, Lendon C, Amouyel P. Is there a relation between APOE expression and brain amyloid load in Alzheimer's disease? J Neurol Neurosurg Psychiatry. 2005; 76:928–33. <u>https://doi.org/10.1136/jnnp.2004.048983</u> PMID:<u>15965197</u>
- 40. Lee TS, Goh L, Chong MS, Chua SM, Chen GB, Feng L, Lim WS, Chan M, Ng TP, Krishnan KR. Downregulation

of TOMM40 expression in the blood of Alzheimer disease subjects compared with matched controls. J Psychiatr Res. 2012; 46:828–30. https://doi.org/10.1016/j.jpsychires.2012.03.006 PMID:22472643

- Nilsen TW. Mechanisms of microRNA-mediated gene regulation in animal cells. Trends Genet. 2007; 23:243–49. <u>https://doi.org/10.1016/j.tig.2007.02.011</u> PMID:<u>17368621</u>
- 42. Kang HS, Kim J, Jang SG, Kwon SY, Park YS, Green JE, Kim HK, Ro J. MicroRNA signature for HER2-positive breast and gastric cancer. Anticancer Res. 2014; 34:3807–10. PMID:24982406
- Yao Y, Suo AL, Li ZF, Liu LY, Tian T, Ni L, Zhang WG, Nan KJ, Song TS, Huang C. MicroRNA profiling of human gastric cancer. Mol Med Rep. 2009; 2:963–70. PMID:21475928
- 44. Li G, Song Y, Li YD, Jie LJ, Wu WY, Li JZ, Zhang Q, Wang Y. Circulating miRNA-302 family members as potential biomarkers for the diagnosis of acute heart failure. Biomark Med. 2018; 12:871–80. https://doi.org/10.2217/bmm-2018-0132 PMID:29900754
- Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. Gigascience. 2015; 4:7. <u>https://doi.org/10.1186/s13742-015-0047-8</u> PMID:<u>25722852</u>
- Prestia A, Caroli A, van der Flier WM, Ossenkoppele R, Van Berckel B, Barkhof F, Teunissen CE, Wall AE, Carter SF, Schöll M, Choo IH, Nordberg A, Scheltens P, Frisoni GB. Prediction of dementia in MCI patients based on core diagnostic markers for Alzheimer disease. Neurology. 2013; 80:1048–56.

https://doi.org/10.1212/WNL.0b013e3182872830 PMID:23390179

- 47. Fischl B, Salat DH, Busa E, Albert M, Dieterich M, Haselgrove C, van der Kouwe A, Killiany R, Kennedy D, Klaveness S, Montillo A, Makris N, Rosen B, Dale AM. Whole brain segmentation: automated labeling of neuroanatomical structures in the human brain. Neuron. 2002; 33:341–55. <u>https://doi.org/10.1016/S0896-6273(02)00569-X</u> PMID:<u>11832223</u>
- Risch N, Merikangas K. The future of genetic studies of complex human diseases. Science. 1996; 273:1516–17. <u>https://doi.org/10.1126/science.273.5281.1516</u> PMID:<u>8801636</u>

SUPPLEMENTARY MATERIALS

Supplementary Figures



Supplementary Figure 1. Multidimensional scaling plot of ADNI samples. Note: Cryptic relatedness and population substructure were checked with genomic identity-by-descent (IBD) and multidimensional scaling (MDS) components. (A) MDS plot of ADNI non-Hispanic Caucasian samples. Samples seemed to form loose clusters and two samples were outliers based on the second MDS component (at top of plot; 031_S_4032 and 031_S_4203), suggesting potential population substructure. To check for cryptic relatedness, which can confound GWAS (genome-wide association study), pairwise identity-by-descent fraction (π) between each pair of samples were calculated using PLINK. Four related sample pairs ($\pi > 0.05$) were identified (031_S_4032 and 031_S_4203 ; 137_S_4466 and 021_S_0159 ; 023_S_0058 and 023_S_4035 ; 024_S_2239 and 024_S_4084), which are probably first-degree relatives. No other cryptic relations were identified from the sample, at a threshold of $\pi > 0.05$. (B) MDS plot of ADNI samples overlaid on HapMap samples. The ancestry of the HapMap participants is shown by the point color. No outlying point was shown. Abbreviations: ADNI, Alzheimer's Disease Neuroimaging Initiative; ASW, African ancestry in Southwest USA; CEU, Utah residents with Northern and Western European ancestry from the CEPH collection; CHB, Han Chinese individuals from Beijing, China; CHD, Chinese in Metropolitan Denver, Colorado; GIH, Gujarati Indians in Houston, Texas; JPT, Tokyo, Japan; LWK, Luhya in Webuye, Kenya; MEX, Mexican ancestry in Los Angeles, California; MKK, Maasai in Kinyawa, Kenya; TSI, Tuscans in Italy; YRI, Yoruba in Ibadan, Nigeria.



Supplementary Figure 2: Manhattan plot for associations with hippocampal atrophy rates after controlling for APOE $\varepsilon 4$. Note: Observed $-\log_{10}$ P-values (y-axis) are shown for all tested single nucleotide polymorphisms on each autosomal chromosome (x-axis) after controlling for age, gender, APOE $\varepsilon 4$ status, education, MRI (magnetic resonance imaging), ICV (intracranial volume) and the first three principal components. No genome-wide significant associations (P < 5 × 10⁻⁸; red line) and suggestive associations (P < 1 × 10⁻⁵; blue line) with hippocampal atrophy rates were identified.



Supplementary Figure 3. Plot of linkage disequilibrium between rs4420638 and rs56131196. R²=0.987, D'=0.993.



Q-Q plot of GWAS p-values

Supplementary Figure 4: Quantile-Quantile plot. Abbreviations: GWAS, genome-wide association study; Q-Q plot, Quantile-Quantile plot.



Supplementary Figure 5. Impact of rs4420638 and rs157582 on cognitive scores and brain structures at baseline. (A) Minimental State Examination (MMSE) score. (B) Alzheimer Disease Assessment Scale-cognitive subscale 11 (ADAS-cog 11) score. (C) Entorhinal volume. (D) Ventricular volume.

Α

Rs4420638

Trans-eQTLs

 P-value
 SNP
 SNP Chr. SNP Chr. position
 Probe Chr. Probe Chr. position
 SNP Alleles Minor Allelez.score Gene name FDR

 1.4287226161643386E-5rs4420638 19
 50114786
 4070039 13
 88664889
 G/A
 G
 4.34
 0.48



Supplementary Figure 6. Bioinformatics analyses. Abbreviations: Chr, chromosome; CRBL, cerebellar cortex; eQTL, expression quantitative trait loci; FCTX, frontal cortex; FDR, false discovery rate; HIPP, hippocampus; log, logarithm; MEDU, medulla; OCTX, occipital cortex; PUTM, putamen; SNIG, substantia nigra; SNP, single nucleotide polymorphism; TCTX, temporal cortex; THAL, thalamus; WHMT, intralobular white matter.

Supplementary Tables

CHR	BP	SNP	MAF	Closest Gene	SNP Type	BETA	Р
19	45395619	rs2075650	G=0.1194	TOMM40	intron	-0.005827	5.11E-08
18	27797293	rs4271662	A=0.3606	MIR302F	intergenic	-0.003789	5.86E-06
18	27801582	rs2900721	T=0.3458	MIR302F	intergenic	-0.00372	8.76E-06

Supplementary Table 1. Suggestive SNPs related to hippocampal atrophy rates without APOE ɛ4 fitted as a covariate.

Abbreviations: BP, base pair (variant position); CHR, chromosome; MAF, minor allele frequency; SNP, single nucleotide polymorphism.

CHR	BP	SNP	MAF	Closest Gene	SNP Type	BETA	Р
18	27797293	rs4271662	A=0.3606	MIR302F	intergenic	-0.003464	1.68E-05
4	66628221	rs4382087	C=0.4427	LOC100144602	intergenic	0.003585	1.96E-05
15	100819272	rs7494886	C=0.1394	ADAMTS17	intron	-0.003804	1.97E-05
11	70002987	rs3781658	A=0.4655	ANO1	intron	-0.003635	2.20E-05
6	157915204	rs2365386	T=0.3984	ZDHHC14	intron	0.003528	2.27E-05
18	27801582	rs2900721	T=0.3458	MIR302F	intergenic	-0.003413	2.28E-05
12	69477770	rs12582988	A=0.1430	СРМ	intergenic	0.004236	2.53E-05
10	81061724	rs10824740	A=0.3472	ZMIZ1	intron	-0.003841	2.62E-05
4	189850773	rs10001577	T=0.4493	LOC285442	intergenic	0.003894	2.74E-05
4	189860582	rs62341079	T=0.4171	LOC285442	intergenic	0.003852	3.08E-05
21	40024830	rs459813	T=0.2865	ERG	intron	0.004252	3.42E-05
3	61834942	rs6766943	G=0.3464	PTPRG	intron	0.004118	3.60E-05
4	96811611	rs4699474	G=0.2482	PDHA2	intergenic	-0.004146	3.62E-05
14	105149313	rs12884142	C=0.1208	MIR4710	intergenic	-0.003959	3.97E-05
4	189862164	rs12647666	T=0.4169	LOC285442	intergenic	0.003785	4.01E-05
6	19824025	rs6921758	T=0.4718	LOC100506885	intergenic	-0.003646	4.20E-05
3	151911863	rs323613	G=0.2316	LOC101928166	intron	-0.00401	4.46E-05
6	31069489	rs9263565	T=0.3165	C6orf15	intergenic	0.003405	4.47E-05
1	4626267	rs241272	G=0.4607	AJAP1	intron	0.003785	4.55E-05
2	139029250	rs6710702	T=0.2165	RPL15P5	intergenic	-0.003783	4.62E-05
19	2430232	rs743578	C=0.3878	LMNB2	intron	-0.00364	4.80E-05
6	112999076	rs4945909	G=0.4718	PA2G4P5	intergenic	0.003288	4.82E-05
15	100807344	rs11634485	T=0.3820	ADAMTS17	intron	-0.003412	4.93E-05
10	81064719	rs2296425	A=0.2648	ZMIZ1	intron	-0.004008	5.00E-05

Supplementary Table 2. Top SNPs related to hippocampal atrophy rates with APOE ɛ4 fitted as a covariate.

Abbreviations: BP, base pair (variant position); CHR, chromosome; MAF, minor allele frequency; SNP, single nucleotide polymorphism.

Supplementary Table 3. COX regression with age and gender as covariates among different genotypes of rs4420638 and rs157582.

Rs4420638							
	Om	nibus Tests of Mo	del Coeffi	cients			
	-2 LOG Likelihood	Over		Change From Previous Block			
Step		Chi-square	df	Sig.	Chi- square	df	Sig.
1	2077.519	47.256	2	0.000	41.495	2	0.000

a. Beginning Block Number 1. Method = Forward Stepwise (Likelihood Ratio)

Variables in the Equation									
	D	SE	Wald	đf	Sia	Exp (B) -	95.0% CI f	for Exp (B)	
	D	SE	vv alu	ui	Sig.		Lower	Upper	
Step 1 group			43.104	2	0.000				
group (1)	1.365	0.234	34.142	1.000	0.000	3.917	2.478	6.191	
group (2)	0.794	0.160	24.504	1.000	0.000	2.212	1.615	3.029	

Note: Group represents AA genotype; group (1) represents GG genotype; group (2) represents AG genotype.

Rs157582								
Omnibus Tests of Model Coefficients								
Step		Over		Change From Previous Block				
	-2 LOG Likelihood	Chi-square	df	Sig.	Chi- square	df	Sig.	
1	1715.267	34.083	2	0.000	26.558	2	0.000	

a. Beginning Block Number 1. Method = Forward Stepwise (Likelihood Ratio)

Variables in the Equation										
	P	SF	Wald	đ£	Sia	Exp (B)	95.0% CI f	for Exp (B)		
	D	SE	vv alu	ui	Sig.		Lower	Upper		
Step 1 group			30.620	2	0.000					
group(1)	1.433	0.263	29.672	1.000	0.000	4.190	2.502	7.016		
group(2)	0.533	0.179	8.911	1.000	0.003	1.704	1.201	2.418		

Note: Group represents CC genotype; group (1) represents TT genotype; group (2) represents CT genotype. Abbreviations: B, beta; CI, confidence interval; df, degrees of freedom; Exp, exponent; LOG, logarithm; SE, standard error; Sig, significance.